



# The reciprocal regulation of stress hormones and GABA<sub>A</sub> receptors

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Stress-derived steroid hormones regulate the expression and function of GABA<sub>A</sub> receptors (GABA<sub>A</sub>Rs). Changes in GABA<sub>A</sub>R subunit expression have been demonstrated under conditions of altered steroid hormone levels, such as stress, as well as following exogenous steroid hormone administration. In addition to the effects of stress-derived steroid hormones on GABA<sub>A</sub>R subunit expression, stress hormones can also be metabolized to neuroactive derivatives which can alter the function of GABA<sub>A</sub>Rs. Neurosteroids allosterically modulate GABA<sub>A</sub>Rs at concentrations comparable to those during stress. In addition to the actions of stress-derived steroid hormones on GABA<sub>A</sub>Rs, GABA<sub>A</sub>Rs reciprocally regulate the production of stress hormones. The stress response is mediated by the hypothalamic-pituitary-adrenal (HPA) axis, the activity of which is governed by corticotropin releasing hormone (CRH) neurons. The activity of CRH neurons is largely controlled by robust GABAergic inhibition. Recently, it has been demonstrated that CRH neurons are regulated by neurosteroid-sensitive, GABA<sub>A</sub>R  $\delta$  subunit-containing receptors representing a novel feedback mechanism onto the HPA axis. Further, it has been demonstrated that neurosteroidogenesis and neurosteroid actions on GABA<sub>A</sub>R  $\delta$  subunit-containing receptors on CRH neurons are necessary to mount the physiological response to stress. Here we review the literature describing the effects of steroid hormones on GABA<sub>A</sub>Rs as well as the importance of GABA<sub>A</sub>Rs in regulating the production of steroid hormones. This review incorporates what we currently know about changes in GABA<sub>A</sub>Rs following stress and the role in HPA axis regulation.

**Keywords: GABA, stress, inhibition, corticosterone, CRH**

GABA<sub>A</sub>Rs are regulated by stress-derived steroid hormones and neurosteroids [for review see Belelli et al. (2009); Maguire and Mody (2009); Gunn et al. (2011)]. Conversely, the HPA axis, and thus the production of stress-derived steroid hormones and neurosteroids, is under robust GABAergic control [for review see Herman et al. (2004); Gunn et al. (2011)].

## GABAERGIC REGULATION OF THE HPA AXIS

Stress induces a physiological response which is mediated by the HPA axis. CRH is released from the hypothalamus and acts in the pituitary to signal the release of adrenocorticotrophic hormone (ACTH), which triggers the release of cortisol from the adrenal gland in humans (corticosterone in mice). The HPA axis is regulated by inputs from numerous different brain regions, involving multiple neurotransmitter systems, as well as the feedback of steroid hormones acting on mineralocorticoid receptors (MRs) and glucocorticoid receptors (GRs) [for review see Herman et al. (2003); Larsen et al. (2003); Ulrich-Lai and Herman (2009)]. These inputs impinge on CRH neurons in the paraventricular nucleus (PVN), which mediate the output of the HPA axis. Although CRH neurons receive a wide variety of inputs from diverse brain regions, their activity is ultimately regulated by GABAergic inhibition [for review see Decavel and van den Pol (1990); Herman et al. (2004)].

A role for GABA in HPA axis regulation has been well established. CRH neurons receive robust GABAergic inhibition (Decavel and van den Pol, 1990, 1992) [for review see Herman et al. (2004); Cullinan et al. (2008)]. It has been suggested that a third of the inputs onto CRH neurons are GABAergic and the density of GABAergic synapses in the parvocellular division of the PVN has been estimated to be above  $20 \times 10^6$  synaptic contacts per mm<sup>3</sup> (Miklos and Kovacs, 2002), highlighting the importance of GABAergic inhibition in the regulation of CRH neurons. In addition, microinjection of GABA antagonists, such as bicuculline, into the PVN activates the HPA axis (Cullinan et al., 2008; Marques de and Franci, 2008) and microinfusion of GABA agonists, such as the stress-derived neurosteroid, THDOC, into the PVN decreases circulating levels of stress hormones (Sarkar et al., 2011).

GABA inputs onto CRH neurons originate primarily from local interneurons surrounding the PVN (peri-PVN) as well as from the subparaventricular zone, the anterior hypothalamic area, dorsomedial hypothalamic nucleus, the medial preoptic area, lateral hypothalamic area, and from multiple nuclei within the bed nucleus of the stria terminalis (BNST) (Cullinan et al., 1993; Roland and Sawchenko, 1993) [for review see Herman et al. (2004); Cullinan et al. (2008)]. In addition to the direct inhibitory connections from these brain regions, CRH neurons also receive

indirect inhibition from other regulatory brain regions including limbic and cortical regions which exert their influences on CRH neurons via interneuron mediators [for review see Herman et al. (2004); Cullinan et al. (2008)].

Despite the well-established role for GABAergic control of the HPA axis at the level of the PVN, very little is known about the GABA<sub>A</sub>R subtypes which mediate the GABAergic control over CRH neurons. GABA<sub>A</sub>Rs are members of the large “Cys-loop” super-family of evolutionarily related and structurally similar ligand-gated ion channels. To-date, 19 different subunits;  $\alpha$ 1-6,  $\beta$ 1-3,  $\gamma$ 1-3,  $\delta$ ,  $\epsilon$ ,  $\theta$ ,  $\pi$ , and  $\rho$ 1-3 have been identified (Barnard et al., 1998; Whiting et al., 1999), which form heteropentameric receptors predominantly composed of 2  $\alpha$ s, 2  $\beta$ s, and either the  $\gamma$ 2 or the  $\delta$  subunit. Depending on their subunit composition, GABA<sub>A</sub>Rs have specific anatomical distributions (Pirker et al., 2000) including subcellular localization (Kittler et al., 2002), kinetics, and pharmacology (Hevers and Luddens, 1998; Mody and Pearce, 2004). GABA<sub>A</sub>Rs mediate two distinct forms of GABAergic inhibition, tonic, and phasic, which are mediated by GABA<sub>A</sub>Rs with unique subunit assemblies (Farrant and Nusser, 2005). Extrasynaptically localized  $\delta$  subunit-containing receptors mediate tonic GABAergic inhibition in many brain regions and confer neurosteroid sensitivity (Mihalek et al., 1999; Belelli et al., 2002; Brown et al., 2002; Wohlfarth et al., 2002; Spigelman et al., 2003). Only recently has it been demonstrated that these neurosteroid-sensitive,  $\delta$  subunit-containing GABA<sub>A</sub>Rs play a pivotal role in the regulation of stress reactivity (Sarkar et al., 2011).

Several GABA<sub>A</sub>R subunits have been identified within the PVN (Fritschy and Mohler, 1995). However, it has been historically difficult to conclusively determine which GABA<sub>A</sub>R subunits are expressed on the CRH neurons within the PVN due to the inability to specifically identify this subset of neurons within this heterogeneous nucleus. Dual hybridization histochemical studies have demonstrated mRNA expression of the GABA<sub>A</sub>R  $\alpha$ 1,  $\alpha$ 2,  $\beta$ 1-3, and  $\gamma$ 1-2 subunits in CRH neurons (Cullinan, 2000). Due to the sparse number of studies that have attempted to identify the specific GABA<sub>A</sub>R subtypes controlling CRH neurons, this list remains incomplete. Information regarding the GABA<sub>A</sub>R subtypes involved in regulation of CRH neurons will provide insight into pharmacological tools which may modulate HPA axis activity. It has recently been demonstrated that rostral ventrolateral medulla (RVLM)-projecting parvocellular neurons in the PVN are regulated by a THIP-sensitive tonic current (Park et al., 2007), indicating that neurosteroid-sensitive, extrasynaptic  $\delta$  subunit-containing GABA<sub>A</sub>Rs may play a role in the regulation of these neurons (Boehm et al., 2006; Mortensen et al., 2010). Further, recent studies have demonstrated GABA<sub>A</sub>R  $\delta$  subunit expression in the PVN and GABA<sub>A</sub>R  $\delta$  subunit-mediated tonic GABAergic control of CRH neurons (Sarkar et al., 2011). These findings demonstrate that GABA<sub>A</sub>R  $\delta$  subunit-containing receptors on CRH neurons play a role in the regulation of the HPA axis.

Stress-derived steroid hormones can be metabolized to neuroactive derivatives, termed neurosteroids, such as the stress-derived neurosteroid,  $3\alpha$ , 21-dihydroxy-5 $\alpha$ -pregnan-20-one (THDOC), and the ovarian-derived neurosteroid,  $3\alpha$ -hydroxy-5 $\alpha$ -pregnan-20-one (allopregnanolone). Neurosteroids

are positive allosteric modulators of GABA<sub>A</sub>Rs (Barker et al., 1986; Majewska et al., 1986; Puia et al., 1990; Purdy et al., 1991; Lambert et al., 1995; Morrow et al., 1995; Hosie et al., 2006; Smith et al., 2007), acting on a neurosteroid binding site identified on GABA<sub>A</sub>Rs (Hosie et al., 2006). It has been demonstrated that neurosteroids act preferentially on GABA<sub>A</sub>R  $\delta$  subunit-containing receptors (Mihalek et al., 1999; Belelli et al., 2002; Brown et al., 2002; Wohlfarth et al., 2002; Spigelman et al., 2003) at physiologically relevant concentrations (Stell et al., 2003). These data are consistent with previous findings demonstrating changes in GABA<sub>A</sub>R  $\delta$  subunit expression in parvocellular neurons in the PVN following stress (Verkuyl et al., 2004), implicating these receptors in the regulation of the stress response. In response to stress, THDOC and allopregnanolone are released at levels which can potentially modulate GABA<sub>A</sub>Rs (Barker et al., 1986; Majewska et al., 1986; Puia et al., 1990; Purdy et al., 1991; Lambert et al., 1995; Morrow et al., 1995; Barbaccia et al., 1996a,b; Hosie et al., 2006; Smith et al., 2007). Under basal conditions, neurosteroids can exert a negative feedback onto the HPA axis, decreasing CRH and ACTH levels (Patchev et al., 1994, 1996) [for review see Morrow (2007)]. Recent data demonstrate a role for neurosteroid actions on GABA<sub>A</sub>R  $\delta$  subunit-containing receptors on CRH neurons in the regulation of the HPA axis (Sarkar et al., 2011), and thus, production of stress hormones. This study demonstrates a decrease in the firing rate of CRH neurons upon the addition of a low concentration of THDOC (10 nM) under basal conditions (Sarkar et al., 2011). Further, the role of the GABA<sub>A</sub>R  $\delta$  subunit in the neurosteroid regulation of CRH neurons was confirmed by demonstrating the loss of this regulation in mice lacking the GABA<sub>A</sub>R  $\delta$  subunit (*Gabrd*<sup>-/-</sup> mice). Together, there is ample evidence that under normal conditions, there is a basal GABAergic inhibition of CRH neurons.

Interestingly, the effects of GABA on CRH neurons are dramatically altered following stress. Stress activates GABAergic neurons which project to the PVN (Cullinan et al., 1995; Campeau and Watson, 1997), which would intuitively suggest inhibition of the HPA axis rather than activation. However, GABA agonists have been shown to increase stress-induced corticosterone levels (Borycz et al., 1992; Sarkar et al., 2011) and blocking production with finasteride has been shown to blunt the corticosterone response to stress (Sarkar et al., 2011). However, due to the fact that both THDOC and allopregnanolone levels are elevated following stress, it isn't clear which of these neurosteroids are responsible for activation of the HPA axis. The role of neurosteroids on GABA<sub>A</sub>R  $\delta$  subunit-containing receptors in the activation of the HPA axis following stress (Sarkar et al., 2011), implicates excitatory actions of GABA in regulation of the HPA axis. Recent evidence suggests that there are deficits in GABAergic control of CRH neurons following stress due to a depolarizing shift in the reversal potential for chloride (Cl<sup>-</sup>) (Hewitt et al., 2009). The inhibitory effects of GABA require the maintenance of the Cl<sup>-</sup> gradient, which is primarily accomplished by the K<sup>+</sup>/Cl<sup>-</sup> co-transporter, KCC2, in the adult brain (Rivera et al., 1999; Payne et al., 2003; Rivera et al., 2005). The surface expression and activity of KCC2 is regulated by phosphorylation of KCC2 residue Ser940 (Lee et al., 2007). Dephosphorylation of KCC2 residue Ser940 and downregulation

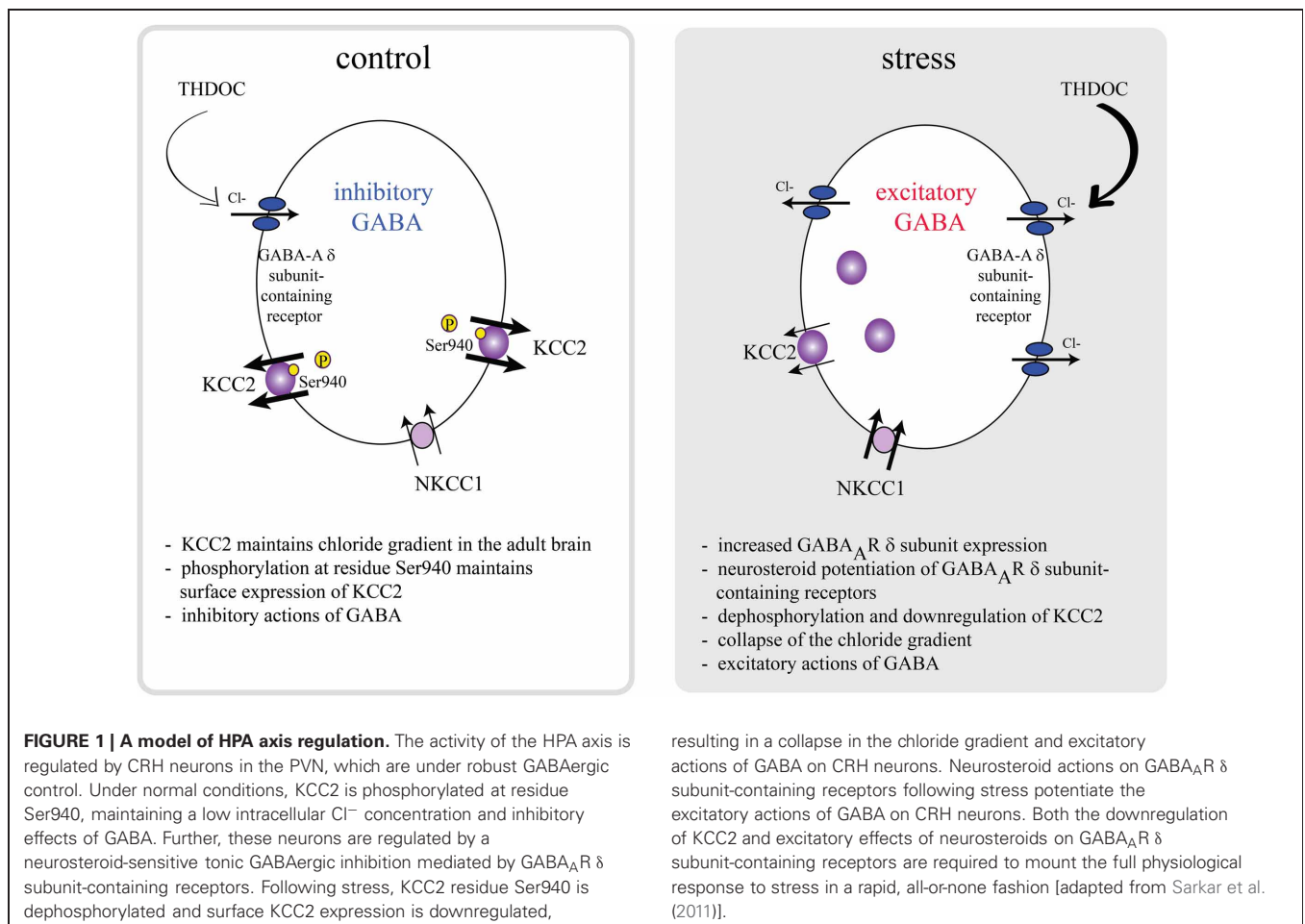
of KCC2 results in depolarizing and excitatory actions of GABA *in vitro* (Lee et al., 2011). Recently, it has been demonstrated that KCC2 plays a role in the regulation of the HPA axis (Sarkar et al., 2011). Following stress, there is a dephosphorylation of KCC2 residue Ser940 and downregulation of surface KCC2 expression in the PVN (Sarkar et al., 2011), resulting in excitatory actions of GABA on CRH neurons (Sarkar et al., 2011). Consistent with excitatory actions of GABA on CRH neurons following stress, THDOC increases the activity of CRH neurons and increases the corticosterone response to stress (Sarkar et al., 2011). The GABA-mediated activation of CRH neurons following acute stress is due to a collapse in the chloride gradient as previously demonstrated (Hewitt et al., 2009) and depolarizing and excitatory actions of GABA (Sarkar et al., 2011), overriding the inhibitory constraint of CRH neurons. These data demonstrate dramatic alterations in GABAergic control of CRH neurons following stress mediated by neurosteroids rather than the actions of steroid hormones on MRs or GRs. We propose a model in which rapid dephosphorylation and downregulation of KCC2 is the most efficient mechanism to overcome the robust GABAergic constraint of CRH neurons to mount a rapid, all-or-none stress response (Figure 1) (Sarkar et al., 2011). This model suggests that both downregulation of KCC2, resulting in excitatory actions of GABA

and neurosteroid potentiation of GABA<sub>A</sub>  $\delta$  subunit-containing receptors is required to mount the full physiological response to stress.

### STRESS HORMONE REGULATION OF GABAERGIC INHIBITION

In addition to the well-established role of GABAergic transmission in the regulation of the HPA axis as outlined above, conversely, stress hormones can also alter GABAergic inhibition. This review will focus on changes that occur in adulthood and will not discuss the vast literature documenting changes in GABAergic inhibition resulting from early life stress. For a more in-depth review of the role of neurosteroids in stress, including prenatal stress, see (Gunn et al., 2011).

Acute and chronic stress has been shown to alter the expression of both GAD and GABA (Yoneda et al., 1983; Otero Losada, 1988; Maroulakou and Stylianopoulou, 1991; Acosta et al., 1993; Bowers et al., 1998) [for review see Cullinan et al. (2008)]. Increased GAD65 and GAD67 expression have been demonstrated following stress in brain regions associated with the regulation of the HPA axis, including the anterior hypothalamic area, dorsomedial nucleus, medial preoptic area, suprachiasmatic nucleus, anterior BST, perifornical nucleus, and peri-PVN region [Bowers et al., 1998; for review see Cullinan et al. (2008)].



resulting in a collapse in the chloride gradient and excitatory actions of GABA on CRH neurons. Neurosteroid actions on GABA<sub>A</sub>  $\delta$  subunit-containing receptors following stress potentiate the excitatory actions of GABA on CRH neurons. Both the downregulation of KCC2 and excitatory effects of neurosteroids on GABA<sub>A</sub>  $\delta$  subunit-containing receptors are required to mount the full physiological response to stress in a rapid, all-or-none fashion [adapted from Sarkar et al. (2011)].

Despite the upregulation of enzymes responsible for GABA synthesis, the frequency of spontaneous inhibitory postsynaptic currents (sIPSCs) has been shown to be decreased following stress (Verkuyl et al., 2004). Similarly, a high dose of exogenous corticosterone has been shown to decrease mIPSC frequency (Verkuyl et al., 2005) and adrenalectomy increases miniature inhibitory postsynaptic currents (mIPSC) frequency (Verkuyl and Joels, 2003) and the number of GABAergic synapses on CRH neurons (Miklos and Kovacs, 2002). Further, demonstrating presynaptic changes in GABAergic inhibition following stress, the expression of receptors for stress-derived steroid hormones (MRs and GRs) have been identified on GABAergic interneurons in the peri-PVN region and stress hormones have been shown to increase the burst firing of these neurons (Shin et al., 2011). These findings are in contrast with the decreased frequency of both mIPSCs and sIPSCs following stress (Verkuyl et al., 2004) and may represent a compensatory change to restore inhibition in this region following stress. In addition to potential changes in presynaptic GABAergic release suggested by changes in GAD expression and GABA levels, there is also abundant evidence of postsynaptic changes in GABA<sub>A</sub>R subunit expression associated with stress.

There is reduced [<sup>3</sup>H]GABA and [<sup>35</sup>S]TBPS binding following stress suggesting alterations in GABA<sub>A</sub> receptor (GABA<sub>A</sub>R) expression (Skerritt et al., 1981; Schwartz et al., 1987; Akinci and Johnston, 1993; Serra et al., 2000) [for review see Skilbeck et al. (2010)]. One thing is for certain, the changes in binding to GABA<sub>A</sub>Rs following stress is extremely variable and results differ according to gender, paradigm used, and laboratory where the experiments were conducted. These results leave little certainty regarding changes in radio-labeled ligand binding to GABA<sub>A</sub>Rs following stress. Pharmacological changes more consistently point to alterations in GABA<sub>A</sub>R expression following stress. For example, stress and adrenalectomy have both been shown to alter benzodiazepine binding (Majewska et al., 1985; De Souza et al., 1986; Goeders et al., 1986; Miller et al., 1987, 1988; Weizman et al., 1990; Smith et al., 1992). However complex, these data suggest that there are changes in GABA<sub>A</sub>R expression associated with stress.

Studies investigating changes in GABA<sub>A</sub>R subunit expression following stress have demonstrated specific changes in GABA<sub>A</sub>R subtypes. There are brain region-specific alterations in GABA<sub>A</sub>R subunit expression following stress, including decreased GABA<sub>A</sub>R β1 and β2 subunit expression in the PVN following stress, with no change in GABA<sub>A</sub>R α1, α3, γ1, or γ2 expression (Verkuyl et al., 2004). Consistent with a role of extrasynaptic GABA<sub>A</sub>Rs in the regulation of the HPA axis, a significant increase in GABA<sub>A</sub>R α5 subunit expression and a decrease in GABA<sub>A</sub>R δ subunit expression have been demonstrated in the PVN following stress (Verkuyl et al., 2004). In the hippocampus,

GABA<sub>A</sub>R β1 and β2 subunit expression is increased (Cullinan and Wolfe, 2000) and GABA<sub>A</sub>R γ2 subunit expression is decreased (Maguire and Mody, 2007). Increased expression of the predominantly extrasynaptic GABA<sub>A</sub>R δ subunit was demonstrated in the hippocampus following stress (Maguire and Mody, 2007) [for review see Belelli et al. (2009); Maguire and Mody (2009)] and these changes can be mimicked by treatment with THDOC (Maguire and Mody, 2007). Although the exact mechanisms underlying alterations in GABA<sub>A</sub>R subunit expression associated with stress are not fully understood, it is thought that these changes are mediated by the actions of stress hormones and/or stress-derived neurosteroids.

Both steroid hormones and neurosteroids are elevated in response to acute stress (Majewska et al., 1985; Purdy et al., 1991; Barbaccia et al., 1996a,b). Acute stress induces an elevation in circulating levels of THDOC from 1–5 nM to 15–30 nM (Reddy and Rogawski, 2002) [for review see Reddy (2003)]. Stress can increase neurosteroid levels to concentrations which can act directly on GABA<sub>A</sub>Rs to both potentiate the effects of GABA (Purdy et al., 1991; Barbaccia et al., 1996b) as well as alter GABA<sub>A</sub>R subunit expression (Maguire and Mody, 2007). Neurosteroids can potentiate the tonic component of GABAergic inhibition via action on GABA<sub>A</sub>R δ subunit-containing receptors at low concentrations (Stell et al., 2003), can potentiate the phasic component of GABAergic inhibition at higher concentrations, and at very high concentrations have even been shown to directly gate the receptor [for review see Lambert et al. (2009)]. In addition to the potentiation of GABAergic transmission by neurosteroids, steroid hormones themselves can alter synaptic GABAergic transmission (Maggio and Segal, 2009). Corticosterone alters the frequency of spontaneous sIPSCs in the hippocampus via actions on MRs (Maggio and Segal, 2009) and increases the amplitude of sIPSCs via actions on GRs (Maggio and Segal, 2009). Neurosteroidogenesis has been demonstrated to be essential for steroid hormone-linked alterations in GABA<sub>A</sub>R subunit expression (Maguire and Mody, 2007). These alterations in GABA<sub>A</sub>R subunit expression following stress are likely mediated by neurosteroid-mediated effects on GABA<sub>A</sub>R phosphorylation (Brussaard and Kokksma, 2003), which controls GABA<sub>A</sub>R expression [for review see Kittler and Moss (2003)]. These data demonstrate the complex actions of both steroid hormones and neurosteroids on GABA<sub>A</sub>Rs via direct modulation or by altering receptor expression.

The findings highlighted in this review demonstrate a reciprocal regulation of stress hormones and GABA receptors, in that GABAergic transmission plays a key role in the regulation of the HPA axis and the production of stress hormones and stress-derived neurosteroids can alter GABA<sub>A</sub>R subunit expression as well as directly modulate GABAergic transmission.

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